

Resource Summary Report

Generated by [dkNET](#) on Aug 30, 2026

Blue Brain Cell Atlas

RRID:SCR_019266

Type: Tool

Proper Citation

Blue Brain Cell Atlas (RRID:SCR_019266)

Resource Information

URL: <https://bbp.epfl.ch/nexus/cell-atlas/>

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Description: Describes number, types, and positions of cells in all areas of mouse brain. Provides densities and positions of all excitatory, inhibitory and neuromodulatory neurons, as well as astrocytes, oligodendrocytes and microglia in each of brain regions defined in Allen Mouse Brain Atlas. Users can download cell numbers for statistical analysis, cell positions and types for modeling and visualizing brain areas. Underlying workflow uses imaging data from Allen Institute Common Coordinate Framework to generate cell positions and assign their type using API for data access.

Synonyms: The Blue Brain Cell Atlas

Resource Type: atlas, data or information resource

Keywords: Cell number, cell type, cell position, brain area, mouse brain, neuromodulatory neurons, astrocytes, oligodendrocytes, microglia, imaging data, Allen Institute Common Coordinate Framework

Availability: Free, Freely available

Resource Name: Blue Brain Cell Atlas

Resource ID: SCR_019266

Record Creation Time: 20220129T080344+0000

Record Last Update: 20260829T183047+0000

Ratings and Alerts

No rating or validation information has been found for Blue Brain Cell Atlas.

No alerts have been found for Blue Brain Cell Atlas.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 11 mentions in open access literature.

Listed below are recent publications. The full list is available at [dkNET](#) .

Bueckle A, et al. (2026) Cell Type Populations for 3D Anatomical Structures of the Human Reference Atlas. Scientific data, 13(1).

Rybarikova M, et al. (2025) Gene editing for Spinocerebellar ataxia type 3 taking advantage of the human ATXN3L paralog as replacement gene. Gene therapy.

Sridhar S, et al. (2025) Targeting TREM2 signaling shows limited impact on cerebrovascular calcification. Life science alliance, 8(1).

Roussel Y, et al. (2023) Mapping of morpho-electric features to molecular identity of cortical inhibitory neurons. PLoS computational biology, 19(1), e1010058.

Iavarone E, et al. (2023) Thalamic control of sensory processing and spindles in a biophysical somatosensory thalamoreticular circuit model of wakefulness and sleep. Cell reports, 42(3), 112200.

Gandolfi D, et al. (2023) Full-scale scaffold model of the human hippocampus CA1 area. Nature computational science, 3(3), 264.

Gandolfi D, et al. (2022) A realistic morpho-anatomical connection strategy for modelling full-scale point-neuron microcircuits. Scientific reports, 12(1), 13864.

Borges FS, et al. (2022) Large-scale biophysically detailed model of somatosensory thalamocortical circuits in NetPyNE. Frontiers in neuroinformatics, 16, 884245.

Rodarie D, et al. (2022) A method to estimate the cellular composition of the mouse brain from heterogeneous datasets. PLoS computational biology, 18(12), e1010739.

Shi J, et al. (2022) MouseNet: A biologically constrained convolutional neural network model for the mouse visual cortex. PLoS computational biology, 18(9), e1010427.

Zhu Z, et al. (2021) A substantia innominata-midbrain circuit controls a general aggressive response. Neuron, 109(9), 1540.