

Resource Summary Report

Generated by [dkNET](#) on Aug 25, 2026

Virginia University School of Medicine Advanced Microscopy Core Facility

RRID:SCR_018736

Type: Tool

Proper Citation

Virginia University School of Medicine Advanced Microscopy Core Facility
(RRID:SCR_018736)

Resource Information

URL: <https://med.virginia.edu/advanced-microscopy-facility/>

Proper Citation: Virginia University School of Medicine Advanced Microscopy Core Facility (RRID:SCR_018736)

Description: Advanced Microscopy Facility is microscopy service and shared instrumentation facility in University of Virginia biomedical research community. Pprovides expertise and instrument access for electron and light microscopy imaging technologies as well as image analysis tools and services for basic scientists and clinical researchers within University of Virginia and across Commonwealth.

Abbreviations: AMF

Synonyms: University of Virginia UVA Advanced Microscopy Facility, UVA Advanced Microscopy Facility

Resource Type: access service resource, core facility, service resource

Keywords: USEDit, microscopy, electron microscopy, light microscopy, imaging technology, image analysis, training, sample preparation, consultation, ABRF, ABRF

Availability: Open

Resource Name: Virginia University School of Medicine Advanced Microscopy Core Facility

Resource ID: SCR_018736

Alternate IDs: ABRF_1018, SCR_018744

Alternate URLs: <https://coremarketplace.org/?FacilityID=1018>,
<https://med.virginia.edu/advanced-microscopy-facility/services/>

Record Creation Time: 20220129T080341+0000

Record Last Update: 20260821T194151+0000

Ratings and Alerts

No rating or validation information has been found for Virginia University School of Medicine Advanced Microscopy Core Facility.

No alerts have been found for Virginia University School of Medicine Advanced Microscopy Core Facility.

Data and Source Information

Source: [SciCrunch Registry](#)

Usage and Citation Metrics

We found 22 mentions in open access literature.

Listed below are recent publications. The full list is available at [dkNET](#) .

Genetta TL, et al. (2026) ZEB1 Promotes Alternate Lengthening of Telomeres at Multiple Levels. *Cancers*, 18(3).

De Hoyos HN, et al. (2026) ATP8B1-TMEM30B Flippase Activity Maintains Stereocilia Lipid Asymmetry Required for Hearing. *bioRxiv : the preprint server for biology*.

Li S, et al. (2026) A regulatory axis for tonotopic MYO7A expression in cochlear hair cells. *Proceedings of the National Academy of Sciences of the United States of America*, 123(15), e2519742123.

Leonard-Duke J, et al. (2025) Mechanically Tunable Poly(Ethylene Glycol) Diacrylate Hydrogels Reveal Stiffness-Related Impairments in Capillary Sprouting in Experimental Lung Fibrosis. *Microcirculation (New York, N.Y. : 1994)*, 32(5), e70018.

Yadav N, et al. (2025) Synergistic activity of simvastatin and irinotecan chemotherapy against glioblastoma converges on TGF-?? signaling. *Journal of neuro-oncology*, 174(3), 621.

Aragon JW, et al. (2025) Emergence of endothelial subtypes and role of cell cycle control in arterial-venous specification during embryonic vascular development. *Cell reports*, 44(10), 116368.

Daboin C, et al. (2025) Mechanical maturation of human dermal fibroblast-laden microporous annealed particle scaffolds during long-term in vitro culture. *Acta biomaterialia*, 207, 353.

Barbeau MC, et al. (2025) The kinase ERK plays a conserved dominant role in the heterogeneity of epithelial-mesenchymal transition in pancreatic cancer cells. *Science signaling*, 18(899), eads7002.

Rao VG, et al. (2025) Mechanisms of cilia regeneration in *Xenopus* multiciliated epithelium in vivo. *EMBO reports*, 26(8), 2192.

Zatorski JM, et al. (2024) Measurement of Covalent Bond Formation in Light-Curing Hydrogels Predicts Physical Stability under Flow. *Analytical chemistry*, 96(50), 19880.

Yamaguchi M, et al. (2024) Transformation of the Kidney into a Pathological Neuro-Immune-Endocrine Organ. *Circulation research*, 135(10), 1025.

Zeng J, et al. (2024) Protocol for integrating tissue clearing and light-sheet imaging to analyze cancer initiation in mosaic analysis with double markers mouse models. *STAR protocols*, 5(2), 103092.

Cox BP, et al. (2024) Local, Quantitative Morphometry of Fibroproliferative Lung Injury Using Laminin. *American journal of respiratory cell and molecular biology*, 71(1), 23.

Zhao XY, et al. (2024) iNOS is necessary for GBP-mediated *T. gondii* clearance in murine macrophages via vacuole nitration and intravacuolar network collapse. *Nature communications*, 15(1), 2698.

Medrano S, et al. (2024) An efficient inducible model for the control of gene expression in renin cells. *American journal of physiology. Renal physiology*, 327(3), F489.

Zatorski JM, et al. (2024) Measurement of covalent bond formation in light-curing hydrogels predicts physical stability under flow. *bioRxiv : the preprint server for biology*.

Przanowski P, et al. (2023) ANKLE1 cleaves mitochondrial DNA and contributes to cancer risk by promoting apoptosis resistance and metabolic dysregulation. *Communications biology*, 6(1), 231.

Zhao XY, et al. (2023) Inducible nitric oxide synthase (iNOS) is necessary for GBP-mediated *T. gondii* restriction in murine macrophages via vacuole nitration and intravacuolar network collapse. *bioRxiv : the preprint server for biology*.

Cox BP, et al. (2023) Local, Quantitative Morphometry of Fibroproliferative Lung Injury using Laminin. *bioRxiv : the preprint server for biology*.

Karanja F, et al. (2022) Ecdysone exerts biphasic control of regenerative signaling, coordinating the completion of regeneration with developmental progression. *Proceedings of the National Academy of Sciences of the United States of America*, 119(5).