Resource Summary Report

Generated by dkNET on Aug 16, 2026

AAVS1 SA-2A-puro-pA donor

RRID:Addgene_22075
Type: Plasmid

Proper Citation

RRID:Addgene_22075

Plasmid Information

URL: http://www.addgene.org/22075

Proper Citation: RRID:Addgene_22075

Insert Name: SA-Puro

Organism: Homo sapiens

Bacterial Resistance: Ampicillin

Defining Citation: PMID:19680244

Vector Backbone Description: Backbone Size:5890; Vector Backbone:N/A; Vector Types:ZFN donor; Bacterial Resistance:Ampicillin

Comments: Please note that this plasmid contains a non-functional ccdB gene. The use of ccdB survival cells may be needed when subcloning or modifying this vector if the number of transformants is low.

Plasmid Name: AAVS1 SA-2A-puro-pA donor

Record Creation Time: 20220422T222058+0000

Record Last Update: 20260815T081208+0000

Ratings and Alerts

No rating or validation information has been found for AAVS1 SA-2A-puro-pA donor.

No alerts have been found for AAVS1 SA-2A-puro-pA donor.

Data and Source Information

Source: Addgene

Usage and Citation Metrics

We found 20 mentions in open access literature.

Listed below are recent publications. The full list is available at dkNET .

de Lima Balico L, et al. (2025) Genomic insertion of ancestral uricase into human liver cells to determine metabolic consequences of pseudogenization. Scientific reports, 15(1), 26093.

Rath P, et al. (2025) Optimizing approaches for targeted integration of transgenic cassettes by integrase-mediated cassette exchange in mouse and human stem cells. Stem cells (Dayton, Ohio), 43(1).

Nakamura J, et al. (2025) Development of programmable RNA imaging with RNA-guided GFP via click chemistry. Nucleic acids research, 53(20).

Bouchareb A, et al. (2024) Increasing Knockin Efficiency in Mouse Zygotes by Transient Hypothermia. The CRISPR journal, 7(2), 111.

Mariano NC, et al. (2023) Inducible Protein Degradation as a Strategy to Identify Phosphoprotein Phosphatase 6 Substrates in RAS-Mutant Colorectal Cancer Cells. Molecular & cellular proteomics : MCP, 22(8), 100614.

Furuno K, et al. (2022) Gelatin nanofiber mats with Lipofectamine/plasmid DNA complexes for in vitro genome editing. Colloids and surfaces. B, Biointerfaces, 216, 112561.

Hanss Z, et al. (2021) Mitochondrial and Clearance Impairment in p.D620N VPS35 Patient-Derived Neurons. Movement disorders : official journal of the Movement Disorder Society, 36(3), 704.

Dinesh RK, et al. (2020) Transcription factor binding at Ig enhancers is linked to somatic hypermutation targeting. European journal of immunology, 50(3), 380.

Arias-Fuenzalida J, et al. (2019) Automated high-throughput high-content autophagy and mitophagy analysis platform. Scientific reports, 9(1), 9455.

Li P, et al. (2018) Efficiency and Specificity of Targeted Integration Mediated by the Adeno-Associated Virus Serotype 2 Rep 78 Protein. Human gene therapy methods, 29(3), 135.

Kim SI, et al. (2018) Microhomology-assisted scarless genome editing in human iPSCs. Nature communications, 9(1), 939.

Sim X, et al. (2016) A Doxycycline-Inducible System for Genetic Correction of iPSC Disease Models. Methods in molecular biology (Clifton, N.J.), 1353, 13.

Casta??o J, et al. (2016) Expression of MLL-AF4 or AF4-MLL fusions does not impact the efficiency of DNA damage repair. Oncotarget, 7(21), 30440.

Guye P, et al. (2016) Genetically engineering self-organization of human pluripotent stem cells into a liver bud-like tissue using Gata6. Nature communications, 7, 10243.

Sivalingam J, et al. (2016) Multidimensional Genome-wide Analyses Show Accurate FVIII Integration by ZFN in Primary Human Cells. Molecular therapy : the journal of the American Society of Gene Therapy, 24(3), 607.

Oceguera-Yanez F, et al. (2016) Engineering the AAVS1 locus for consistent and scalable transgene expression in human iPSCs and their differentiated derivatives. Methods (San Diego, Calif.), 101, 43.

Gossai NP, et al. (2016) Drug conjugated nanoparticles activated by cancer cell specific mRNA. Oncotarget, 7(25), 38243.

Bobis-Wozowicz S, et al. (2014) Non-integrating gamma-retroviral vectors as a versatile tool for transient zinc-finger nuclease delivery. Scientific reports, 4, 4656.

Xu L, et al. (2013) Targeted Myostatin Gene Editing in Multiple Mammalian Species Directed by a Single Pair of TALE Nucleases. Molecular therapy. Nucleic acids, 2(7), e112.

Dreyer AK, et al. (2012) Zinc-finger nucleases-based genome engineering to generate isogenic human cell lines. Methods in molecular biology (Clifton, N.J.), 813, 145.