

Resource Summary Report

Generated by [dkNET](#) on Sep 18, 2026

pUS250

RRID:Addgene_198322

Type: Plasmid

Proper Citation

RRID:Addgene_198322

Plasmid Information

URL: <http://www.addgene.org/198322>

Proper Citation: RRID:Addgene_198322

Bacterial Resistance: Kanamycin

Vector Backbone Description: Backbone Marker:Coleman; Backbone Size:4690; Vector Backbone:pUS250; Vector Types:Bacterial Expression, Synthetic Biology; Bacterial Resistance:Kanamycin

Comments: CLONING: The multiple cloning site (MCS) in pUS250 functions differently to a typical MCS. You need to choose one unique restriction site on the left side of the amilCP marker gene (e.g. EcoRI), and one unique restriction site on the right side (e.g. PstI) in order for the blue/white selection and the cumate-inducible expression features to work properly. Cutting the plasmid in this way excises the amilCP gene, replacing it with your gene of interest, and changing the phenotype from blue to white. The vector is also compatible with GoldenGate cloning (using either BsaI or Esp3I) and BioBrick cloning (iGEM). In the case of GoldenGate cloning, you don't need to use two different enzymes, you just use one or the other, since each enzyme has two sites, one on each side of amilCP, and each yields a different overhang. HOST RANGE and EXPRESSION: we have shown that pUS250 can be used for cumate-inducible gene expression in E.coli, Pseudomonas putida, and Rhizobium leguminosarum. It is likely that the plasmid will also be useful in other gram negatives (Proteobacteria) since the pBBR replicon has a very broad host range. The plasmid can be transferred by conjugation from E.coli strains such as S17 or SM10 into other species due to the oriT sequence. For the E.coli and Rhizobium, 100 uM cumate is sufficient for expression. For Pseudomonas, this needs to be increased to 10 mM (we think there is an efflux pump that pumps cumate back out of these cells). The cumate should be added after autoclaving. We make a 0.5 M cumate stock solution by mixing equal parts of 1M aqueous Tris base with 1M cumic acid in ethanol. Cumate is an excellent inducer since it is both cheap and non-toxic to both bacteria and people. You can even use cumin (the spice) for induction of gene expression in this plasmid! (there is enough cumate in the cumin). STABILITY and COPY NUMBER:

We estimate that the vector has a copy number of 5-10 in E.coli, so it is certainly a low copy vector. Best to make large-scale plasmid preps (e.g. 50 ml culture) rather than small-scale, in order to ensure you get enough plasmid DNA to work with. The plasmid is quite stable but we have seen white mutants appear occasionally which have deletions in amilCP.

Plasmid Name: pUS250

Record Creation Time: 20230414T080545+0000

Record Last Update: 20260912T093739+0000

Ratings and Alerts

No rating or validation information has been found for pUS250.

No alerts have been found for pUS250.

Data and Source Information

Source: [Addgene](#)

Usage and Citation Metrics

We found 2 mentions in open access literature.

Listed below are recent publications. The full list is available at [dkNET](#) .

Chang C, et al. (2024) Modified Tn7 transposon vectors for controlled chromosomal gene expression. Applied and environmental microbiology, 90(10), e0155624.

Lian ZJ, et al. (2023) Genetic basis of I-complex plasmid stability and conjugation. PLoS genetics, 19(6), e1010773.