Resource Summary Report

Generated by dkNET on Sep 8, 2026

pLVCT-tTR-KRAB

RRID:Addgene_11643
Type: Plasmid

Proper Citation

RRID:Addgene_11643

Plasmid Information

URL: http://www.addgene.org/11643

Proper Citation: RRID:Addgene_11643

Insert Name: CAG promoters, GFP, tTR-KRAB, Tet-on

Bacterial Resistance: Ampicillin

Defining Citation: PMID:16432520

Vector Backbone Description: Backbone Size:12877; Vector Backbone:None; Vector Types:Mammalian Expression, Lentiviral; Bacterial Resistance:Ampicillin

Comments: Transgenes can be expressed from any RNA Pol II promoter as part of bicistronic unit comprising the KRAB-based repressor; tetO sequences are inserted into the vector LTR. Tet-on and Tet-off versions rely on repressors that bind in the absence or the presence of doxycycline, respectively. Addition of Pol III promoter-small hairpin RNA cassette allows for drug-controllable RNA interference (Tet-on shRNA). Cloning shRNA into pLVET, pLVCT, and pLVPT vectors: first clone shRNA into pLVTHM downstream of the tetO-H1 region. Then cut pLVTHM containing your shRNA with MscI-FspI and clone the insert containing the 3'LTR to target plasmid opened with MscI-FspI. FspI cuts into AmpR. This means for inverted clones AmpR will not be restored; after selection you will be left with clones with the shRNA cassette and functional AmpR. pLVTHM and packaging plasmids for this system are also available at Addgene http://www.addgene.org/rnaitools Please note that the full sequence for this plasmid is approximated and not fully verified. Please visit the Trono lab website http://tronolab.epfl.ch to see frequently asked questions on cloning strategies and packaging.

Plasmid Name: pLVCT-tTR-KRAB

Record Creation Time: 20220422T221610+0000

Record Last Update: 20260905T074512+0000

Ratings and Alerts

No rating or validation information has been found for pLVCT-tTR-KRAB.

No alerts have been found for pLVCT-tTR-KRAB.

Data and Source Information

Source: Addgene

Usage and Citation Metrics

We found 17 mentions in open access literature.

Listed below are recent publications. The full list is available at dkNET .

Yamauchi K, et al. (2020) Overexpression of Nuclear Receptor 5A1 Induces and Maintains an Intermediate State of Conversion between Primed and Naive Pluripotency. Stem cell reports, 14(3), 506.

Zhou Q, et al. (2020) Targeting CLK3 inhibits the progression of cholangiocarcinoma by reprogramming nucleotide metabolism. The Journal of experimental medicine, 217(8).

Akimov V, et al. (2018) StUbEx PLUS-A Modified Stable Tagged Ubiquitin Exchange System for Peptide Level Purification and In-Depth Mapping of Ubiquitination Sites. Journal of proteome research, 17(1), 296.

Chen X, et al. (2016) Probing the impact of chromatin conformation on genome editing tools. Nucleic acids research, 44(13), 6482.

Br??gger V, et al. (2015) HDAC1/2-Dependent P0 Expression Maintains Paranodal and Nodal Integrity Independently of Myelin Stability through Interactions with Neurofascins. PLoS biology, 13(9), e1002258.

Li J, et al. (2015) OTX2 is a therapeutic target for retinoblastoma and may function as a common factor between C-MYC, CRX, and phosphorylated RB pathways. International journal of oncology, 47(5), 1703.

Akimov V, et al. (2014) StUbEx: Stable tagged ubiquitin exchange system for the global investigation of cellular ubiquitination. Journal of proteome research, 13(9), 4192.

Solari V, et al. (2014) MYCN-dependent expression of sulfatase-2 regulates neuroblastoma cell survival. Cancer research, 74(21), 5999.

Cai X, et al. (2014) Autonomous stimulation of cancer cell plasticity by the human NKG2D lymphocyte receptor coexpressed with its ligands on cancer cells. PloS one, 9(10), e108942.

Du ZW, et al. (2013) miR-200 and miR-96 families repress neural induction from human embryonic stem cells. Development (Cambridge, England), 140(12), 2611.

Doherty JE, et al. (2013) An adaptable system for improving transposon-based gene expression in vivo via transient transgene repression. FASEB journal : official publication of the Federation of American Societies for Experimental Biology, 27(9), 3753.

Saunders TL, et al. (2011) Inducible transgenic mouse models. Methods in molecular biology (Clifton, N.J.), 693, 103.

Kesireddy V, et al. (2010) Multipurpose modular lentiviral vectors for RNA interference and transgene expression. Molecular biology reports, 37(6), 2863.

Du ZW, et al. (2010) Lentiviral vector-mediated transgenesis in human embryonic stem cells. Methods in molecular biology (Clifton, N.J.), 614, 127.

Du ZW, et al. (2009) Cre recombination-mediated cassette exchange for building versatile transgenic human embryonic stem cells lines. Stem cells (Dayton, Ohio), 27(5), 1032.

Kedinger V, et al. (2009) p110 CUX1 homeodomain protein stimulates cell migration and invasion in part through a regulatory cascade culminating in the repression of E-cadherin and occludin. The Journal of biological chemistry, 284(40), 27701.

Cadieux C, et al. (2009) Mouse mammary tumor virus p75 and p110 CUX1 transgenic mice develop mammary tumors of various histologic types. Cancer research, 69(18), 7188.